Field-Based Plot Extraction Using UAV RGB Images

Changye Yang¹

Sriram Baireddy¹

Melba Crawford²

Edward J. Delp¹

Envu Cai¹

¹Video and Image Processing Laboratory (VIPER), School of Electrical and Computer Engineering

²School of Civil Engineering

Purdue University

West Lafayette, Indiana, USA

Abstract

Unmanned Aerial Vehicles (UAVs) have become popular for use in plant phenotyping of field based crops, such as maize and sorghum, due to their ability to acquire high resolution data over field trials. Field experiments, which may comprise thousands of plants, are planted according to experimental designs to evaluate varieties or management practices. For many types of phenotyping analysis, we examine smaller groups of plants known as "plots." In this paper, we propose a new plot extraction method that will segment a UAV image into plots. We will demonstrate that our method achieves higher plot extraction accuracy than existing approaches.

1. Introduction

Plant phenotyping refers to the characterization and quantification of physical traits of plants such as height, leaf area, biomass, or flowering time [6]. Traditional phenotyping methods involve labor intensive field work [2]. Modern high-throughput methods such as the use of Unmanned Aerial Vehicle (UAV) imaging [16] can drastically reduce the workload and cost [18]. Recent technological improvements in imaging sensors, UAV platforms, and computational hardware also makes UAV systems more accessible [1]. UAV-based imaging systems have been shown to work well for plant phenotyping tasks such as biomass prediction [12], salinity stress analysis [4], and disease detection [15].

Images acquired from UAVs are typically processed to produce geometrically corrected, georeferenced orthomosaic images over an extended area [3, 7]. Figure 1 shows an RGB orthomosiac image of a field of sorghum [14] acquired at an altitude of 40 meters with spatial resolution of 1 centimeter/pixel. A planted field consists of many smaller groups of plants known as "plots" as shown in Figure 2.



Figure 1: RGB orthomosaic image acquired over sorghum with a resolution of 1cm/pixel

Field trials for plant breeding experiments may comprise hundreds to thousands of plots within a field. Plot scale evaluation of phenotypes requires analysis of the responses within individual plots, which must be extracted. Manual plot extraction is not feasible due to the very large number of plots in a field. Using GPS-guided precision planters, fields are often planted in a "grid" pattern as shown in Figure 2. These planting patterns will aid in plant extraction.

In this paper, we propose a plot extraction method to extract the plots from RGB UAV orthomosaic images. We refer to our method as Comb Function Optimization Plot Extraction (COPE). Our method utilizes a series of "comb" functions to locate the gaps between the plots by optimizing 2 energy functions related to the quantity of plants within a given area. COPE can be used on almost any grid-like planted field (e.g., GPS-guided precision planter) and re-



Figure 2: Enlarged 1cm resolution orthomosaic image with plots visible (note the extracted plot shown in red)

quires no pre-training.

2. Background

Several plot extraction methods have been described in the literature. In [13], Tresch *et al.* proposed a plot extraction method known as EasyMPE, which converts the field plant pixels into an energy map. Each plot is identified by thresholding the energy map. In theory, each plot will have two cutoff lines, one before the plot begins and one after the plot ends. In practice, there might be not be exactly two cutoff lines due to variations in the energy function. This can result in missing or extra plots.

In [5], Khan *et al.* proposed a plot extraction method that starts with a uniform-sized grid of plots. The position of each plot is then adjusted by minimizing the inter-plot energy and maximizing intra-plot energy. This method assumes the plots to be fixed size, which is limiting because this is not true for many fields. In addition, this method fails if there are no plants visible in the plot: for example in Figure 2. Possible causes for a lack of plants can be rain damage or no/late germination.

In [10], Prat proposed a method assuming equal distance between plots. Prat first finds the binary plant segmentation mask from the orthomosaic image by thresholding the H color channel of the HSV color space [11]. The resulting plant segmentation mask is then used to form energy functions that are used to find gaps in the plots. Assuming an equal distance between plots, the boundaries of the plots are then found by optimization of the energy functions. The equal distance assumption for the plots causes the resulting extracted plot boundaries to not always lie in the gaps between plots; this approach then needs time-consuming manual adjustments to be useful. Our proposed method resolves the issues described for the previous methods.

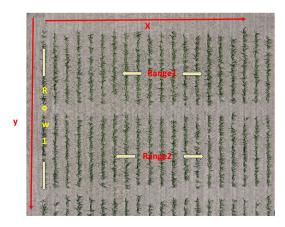


Figure 3: The row and ranges shown for an enlarged orthomosaic image.

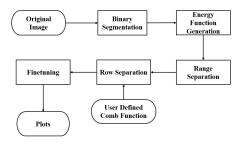


Figure 4: Block diagram of COPE

3. Proposed Method: COPE

We assume that the field has been planted in a grid pattern described above. For convenience, we further assume the field is planted in a north/south orientation as shown in Figure 3. We define a "row" in the orthomosaic image as a north/south (vertical) near-linear group of plots. The "range" is defined to be a east/west (horizontal) near-linear group of plots. The planted field can be divided into rows and ranges as shown in Figure 3. Note a full row in the field will be associated with many plots, and similarly, a range will cross many rows.

For our work, a precision planter drops seeds for a given number of rows (denoted as C) simultaneously. The planter drives the length of the field, and the seeds of a given variety are dropped for the predetermined distance for a plot; no seeds are dropped between the ranges (creating gaps). The planter turns around at the end of the field and plants the next set of C rows.

Figure 4 shows a block diagram of COPE. The plants are segmented from the orthomosaic image to form a binary segmentation mask. The number of plant pixels are counted along the x and y axis in the binary segmentation

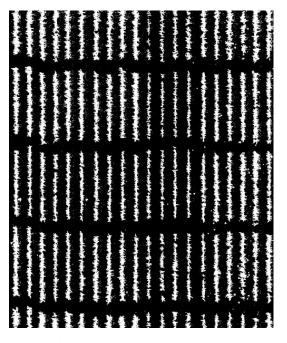


Figure 5: A binary plant segmentation mask with approximately 5 ranges, 20 rows, and 100 plots

mask to form two energy functions described below. Since there are fewer plant pixels in the gaps between the plots, and the rows and ranges are relatively well aligned, the local minima of the energy functions correspond to the gaps between the plots and can be used to define the boundaries of the plots.

3.1. Energy Function and Range Separation

The first step is to segment the plant pixels from the orthomosaic image O(x, y, z) to form the plant segmentation mask I(x, y). A region of interest is extracted from the RGB orthomosaic image and converted to the HSV color space [10, 11]. The H color channel of the orthomosaic image is denoted as the image $O_H(x, y)$. The binary plant segmentation mask I(x, y) at x and y is estimated by thresholding the H channel image in an experimentally determined range of pixel values. For the experiments described later, the pixel value range is 20 to 90:

$$I(x,y) = \begin{cases} 1 & \text{if } 20 \le O_H(x,y) \le 90\\ 0 & \text{else} \end{cases}$$
(1)

A sample plant segmentation mask is shown in Figure 5, note this image has approximately 5 ranges, 20 rows, and 100 plots. Note COPE is independent of the type of plant segmentation used. Other plant segmentation methods such as Otsu [8] can be used. In addition, COPE is not limited to RGB orthomosaic images, any orthomosaic image can be used.

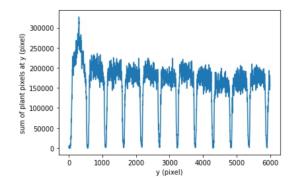


Figure 6: A range energy function

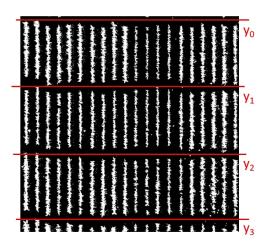


Figure 7: A plant segmentation mask with range separation lines shown in red

We can then estimate the energy functions used for plot extraction. From the plant segmentation mask I(x, y), the range energy function $h_{ra}(y)$ at y and global row energy function $h_{ro-gl}(x)$ at x can be determined by counting the pixels in the plant segmentation mask along the x and y axis:

$$h_{ra}(y) = \sum_{x} I(x, y) \tag{2}$$

$$h_{ro-gl}(x) = \sum_{y} I(x, y) \tag{3}$$

The energy functions can also be thought of as projections or profiles of the x and y spaces. An example of a range energy function $h_{ra}(y)$ is shown in Figure 6. Note the local minima correspond to the gaps between ranges.

Ranges are separated by assuming equal length and distance for all the ranges. Assume the field is defined by Mrows $\times N$ ranges. The range separation lines can be defined as the set of N + 1 horizontal lines that separate each range as shown in red in Figure 7. We assume range separation lines to be Δy apart. We obtain the range separation lines from optimizing the range energy function $h_{ra}(y)$. Therefore we need to find \bar{y}_0 and Δy . Let the set of N + 1 range separation lines be $\{\bar{y}_0, \bar{y}_1, \dots, \bar{y}_N\}$, then

$$\bar{y}_n = \bar{y}_0 + n\Delta y$$

$$\bar{y}_0, \bar{\Delta}y = \arg\min_{y_0, \Delta y} \sum_{n=0}^N h_{ra}(y_0 + n\Delta y)$$
(4)

The equidistant assumption between the range separation lines we made above causes errors in the separation of the ranges due to the start/stopping of the seed drops when planting. Each range separation line is then adjusted to reduce the error. The adjusted range separation lines can be defined as the set $\{\hat{y}_0, \hat{y}_1, \dots, \hat{y}_N\}$, then

$$\hat{y}_{i} = \bar{y}_{i} + \arg\min_{\Delta \bar{y}} h_{ra}(\bar{y}_{i} + \Delta \bar{y}), where$$

$$|\Delta \bar{y}| \le D_{ran-gap}$$
(5)

 $D_{ran-gap}$ is a constraint chosen to restrict the distance correction. For our experiments $D_{ran-gap} \approx 100$.

3.2. Row Separation - Modified Comb Function

With the ranges separated, the local row energy function $h_{ro}^{z}(x)$ of range z at x are obtained:

$$h_{ro}^{z}(x) = \sum_{y=\hat{y}_{i-1}}^{\hat{y}_{i}} I(x, y)$$
(6)

Each range has its own local row energy function.

In order to smooth the optimization process, both the local and global row energy functions are normalized 0 to 1. This results in a modified global row energy function $\hat{h}_{ro-ql}(x)$ and modified local row energy function $\hat{h}_{ro}^{i}(x)$

$$\hat{h}_{ro-gl}(x) = \begin{cases} 1 & \text{if } h_{ro-gl}(x) \ge K \\ & \text{where K is the mean of } h_{rogl}(x) \\ \frac{h_{rogl}(x)}{K} & \text{else} \end{cases}$$
(7)

$$\hat{h}_{ro}^{z}(x) = \begin{cases} 1 & \text{if } h_{ro}^{i}(x) \geq K \\ & \text{where K is the mean of } h_{ro}^{z}(x) & (8) \\ \frac{h_{ro}^{z}(x)}{K} & \text{else} \end{cases}$$

Recall from the previous section, the planter plants C rows of crops at fixed distances apart simultaneously. In each range, we refer to these C rows planted together as a "crop set." An example of crop sets are shown in Figure 8. Recall that M is total number of rows in the range. Each

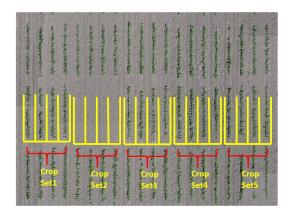


Figure 8: Sets of crops ("crop sets") relative to their range - note the comb-like structure overlaid on the image and shown in yellow

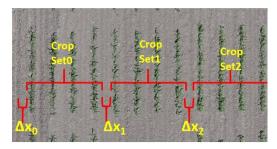


Figure 9: Crop sets offsets relative to their range

range can be represented by joining $\frac{M}{C}$ crop sets together in the range direction as shown in Figure 8. We refer to the distances between each crop set as the "crop set offset" denoted as Δx_i where $0 \le i < \frac{M}{C}$. Sample crop set offsets are shown in Figure 9. Note the positions of the plot boundaries inside any crop set are identical across all crop sets. Even though the planter is GPS guided, the distances between the groups of crop sets vary by a small amount relative to the fixed distance between the individual plots inside the crop set provided by the planter. In order to find the plot boundaries for the range, the only variable required is the crop set offsets.

We need to locate the crop sets by using the energy functions to find the crop set offsets. Since a crop set contains plots with fixed boundaries, the corresponding local minima of the energy functions are a fixed distance apart. In order to find the local minima on the energy functions, a comb function f(m) is formed. The comb function is defined by its width and the number of "comb spikes." Examples of comb functions are shown in yellow in Figure 8 where the combs are 300 pixels wide and there are 5 comb spikes. Let the width of f(m) be the same as the width of a crop set and the number of comb spikes to be C + 1 with the comb

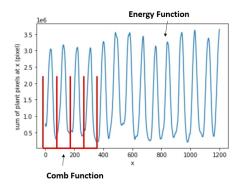


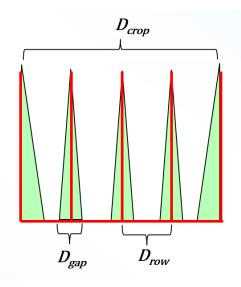
Figure 10: The comb function spikes are fit to the local minima of the energy function

spikes placed in the middle of the gaps between the plots (Figure 8). The parameters of the comb function depends on the shape and dimension of the planting pattern as well as the orthomosaic image resolution. We align the comb function with the energy function such that the comb spikes are at the local minimum as shown in Figure 10. The location of the crop set in the field can then be determined.

Let D_{gap} be the average gap (in pixels) width between plots in the same crop set as shown in Figure 11. D_{gap} is used to modify the comb spikes to restrain the comb spikes to the gap between the plots. The comb function f(m) is altered to form the "modified comb function" $\hat{f}(m)$ by convolving the comb function with a triangle function to widen the comb spikes. The width of the triangle function is the same as the plot gap width D_{gap} and the height is 1. The function is then trimmed to the same width as the crop set to form $\hat{f}(m)$ as shown Figure 11.

Each range can be represented by joining $\frac{M}{C}$ crop sets together side by side, with each crop set having a crop set offset of Δx_i as shown in Figure 9. To find the crop set offsets, starting at range z, we estimate first the crop set offset Δx_0 , where:

$$\Delta x_i^z = \arg \min_{\Delta x_i^z} \omega_0 \frac{\Delta x_i^{z^2}}{D_{row}^2} + \omega_1 loss_1 + \omega_2 loss_2$$
where $loss_1 = \frac{2}{D_{gap}} \hat{f}(n) \cdot \hat{h}_{ro}^z(n)$
 $loss_2 = \frac{2}{D_{gap}} \hat{f}(n) \cdot \hat{h}_{ro-gl}(n)$
 $|\Delta x_i| \le D_{gap}$
 $x_{off}^i + \Delta x_i \le n \le x_{off}^i + \Delta x_i + D_{crop}$
 $x_{off}^i = 0$ when $i = 0$
 x_{off}^i is an intermediate parameter
$$(9)$$





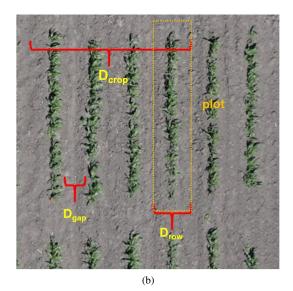


Figure 11: (a) Modified comb function with parameters used in Equation 9 (b) Modified comb function parameters overlaid on the field image

Where D_{row} is the plot width, D_{gap} is the plot gap width, D_{crop} is the crop set width as shown in Figure 11. We then update x_{off}^i by:

$$x_{off}^{i+1} = x_{off}^i + \Delta x_i + D_{crop} \tag{10}$$

After finding Δx_0 , we can find the rest of crop set offsets Δx_i for range z by a similar process.

Crop set boundary lines are defined as the $\frac{M}{C} + 1$ vertical lines that separate the crop sets from each other as shown in Figure 12. Let the set of crop set boundary lines be $\{s_0, s_1, \ldots, s_{\frac{M}{C}}\}$, then:

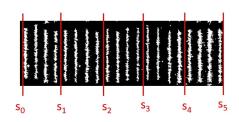


Figure 12: A plant segmentation mask with crop set boundary lines in red

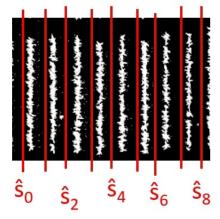


Figure 13: A plant segmentation mask with plot boundary lines in red

$$s_j = \begin{cases} \Delta x_0 & \text{if } j = 0\\ x_{off}^{j-1} + \Delta x_{j-1} + D_{crop} & \text{if } j = \frac{M}{C} \\ \frac{x_{off}^j + \Delta x_j + x_{off}^j}{2} & \text{else} \end{cases}$$
(11)

The plot boundary lines are defined as the M + 1 vertical lines that separate the plots from each other as shown in Figure 13. Let the set of plot boundary lines be $\{\hat{s}_0, \hat{s}_1, \hat{s}_2, \dots, \hat{s}_M\}$, then

$$\hat{s}_m = \begin{cases} s_m & \text{if } m \mod C = 0\\ s_{m-k} + kD_{row} & \text{else}\\ \text{where } k = m \mod C \end{cases}$$
(12)

Using the plot boundary lines and the range separation lines, we can find plot boundaries for any plot in range z. For example if we examine the i_{th} plot in range z: the row boundaries are \hat{s}_{i-1} and \hat{s}_i , the range boundaries are \hat{y}_{z-1} and \hat{y}_z . Repeat the steps above to find the plot boundaries for all the plots across all ranges.



Figure 14: Plots extracted by COPE from a maize orthomosaic image with 0.25cm/pixel resolution. The plots are color coded to make them easier to see

3.3. Boundary Fine-Tuning

At this point all the plots from the same range have the same range boundaries. The final step is to fine tune the range boundaries for each individual plot. Assume a specific plot (i_{th} row and j_{th} range) with range boundaries $y_{top}^{ij}, y_{bot}^{ij}$ and row boundaries $x_{left}^{ij}, x_{right}^{ij}$, the local range energy function $h_{local}^{ij}(y)$ at y is then obtained:

$$h_{local}^{ij}(y) = \begin{cases} \sum_{x} I(x,y) & \text{if } x_{left}^{ij} \le x \le x_{right}^{ij} \text{ and} \\ & y \le y_{top}^{ij} - D_{ran-gap} \text{ and} \\ & y_{bot}^{ij} + D_{ran-gap} \le y \\ 0 & \text{else} \end{cases}$$
(13)

Recall $D_{ran-gap}$ is a constraint chosen to restrict the distance correction. For our experiments $D_{ran-gap} \approx 100$. The adjusted local range energy function $\hat{s}_{ij}(y)$ is normalized using Equation 7. Similar to the modified comb function, a triangle function t(y) with width of $D_{ran-gap}$ and height of 1 is used to assist the optimization (Equation 14). Using the adjusted local range energy function, the finetuned y_{topnew}^{ij} can be estimated by

$$y_{topnew}^{ij} = y_{top}^{ij} + \arg\min_{\Delta y} (\hat{h}_{local}^{ij} * t)(y_{top}^{ij} + \Delta y)$$

$$where |\Delta y| \le D_{ran-gap}$$
(14)

We use Equation 14 to fine tune the range boundaries for all plots.

3.4. Summary of COPE

COPE assumes the field is planted in a grid fashion. Additionally, COPE works better at the early stage of plant growth when the gap between the plants are not closed by the plants. The requirements for COPE are:

- Orthomosaic image
- Plant segmentation mask
- Region of interest from the orthomosaic image
- Number of row M and ranges N in the region of interest
- Range gap width $D_{ran-gap}$
- *C* the number of rows planted at the same time by the planter
- Crop set width D_{crop}
- Plot width D_{row}
- Plot gap width D_{gap}

Note C, D_{crop} , D_{row} , and D_{gap} are parameters used to describe the modified comb function. We only need to describe the modified comb function once unless the planting pattern or image resolution changes. COPE will find the extracted plots, an example is shown in Figure 14.

4. Experimental Results

For our experiments we acquired orthomosaic images of sorghum and maize fields at various dates and altitudes. We evaluated our method (COPE) against EasyMPE [13] and the method described in [10] using our dataset. Note the typical orthomosaic images used in our experiment have more than 300 million pixels (e.g., 11500×47500 pixels). Enlarged regions of an orthomosaic image are shown in Figure 15. From the orthomosaic images, five sub regions (approximately 700 plots) were randomly selected for manual labeling. The manually labeled plot boundaries are used as ground truth for the evaluation of the three plot extraction methods.

The Intersection over Union (IoU) [9] is used to compare the extracted plots. For each of the 3 methods, we estimate the IoU between the extracted plots and the corresponding ground truth.

Methods	IoU
EasyMPE [13]	0.48
Method in [10]	0.65
COPE	0.92

Table 1: Average IoU for different plot extraction methods

The average IoU for each method is shown in Table 1. Our proposed method, COPE, has the highest IoU among

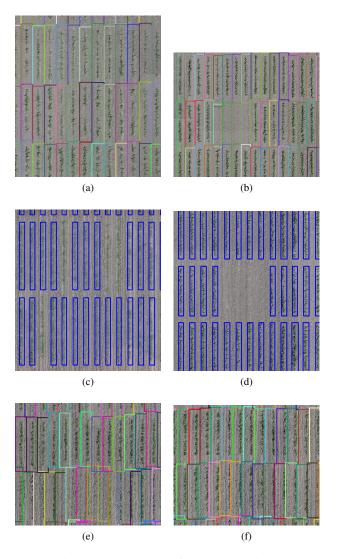


Figure 15: A comparison of plot extraction methods (sorghum image with 1cm/pixel spatial resolution) (a/b) COPE (c/d) EasyMPE [13] (e/f) The method in [10]

all the methods. From Figure 15, it is also visually noted that COPE performs well. EasyMPE [13] has many missing plots and the method in [10] has plot boundaries intersecting with the plant material.

Methods	PBA Time
EasyMPE [13]	1659s
Method in [10]	3712s
COPE	438s

Table 2: Manual plot boundary adjustment (PBA) time inseconds(s) for 700 plots

Manual plot boundary adjustment is sometimes needed

to correct boundary errors of the plot segments in the extracted plots [17]. An experienced post-doctoral researcher, not associated with our team, evaluated each method by estimating the total time required for manual adjustment. For the five sub regions used for ground truthing (700 plots) the plot boundaries found for each of the three methods were manually adjusted by the researcher. The manual plot boundary adjustment (PBA) time for each method for the 700 plots is shown in Table 2. COPE achieves more than a 70% reduction in plot boundary adjustment time compared to the second best method EasyMPE. Note lower manual plot boundary adjustment time indicates many of the extracted plots need no adjustment which corresponds to better automated extraction of the plots.

5. Conclusions and Future Work

Overall COPE is a reliable tool for plot extraction using RGB orthomosaic images. COPE achieves the highest performance in terms of IoU while also requiring the least amount of manual adjustment time. COPE currently requires fields to be planted in a strictly grid-like pattern. In the future, we will improve our method to reduce the reliance on strictly grid-like fields, for example when the the grids are not well aligned in the field. We will also address imagery other than RGB images as well as imagery with different lighting conditions.

The source code for COPE and the datasets used for our experiments will made available to the community. For more information contact Edward J. Delp at ace@ecn.purdue.edu

6. Acknowledgments

We thank Professor Ayman Habib and the Digital Photogrammetry Research Group (DPRG) from the School of Civil Engineering at Purdue University for providing the orthomosaic images used in this paper. We also thank Jieqiong Zhao from Arizona State University for helping with the groundtruthing. The work presented here was funded by the Advanced Research Projects Agency-Energy (ARPA-E), U.S. Department of Energy, under Award Number DE-AR0001135. The views and opinions of the authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof. Address all correspondence to Edward J. Delp, ace@ecn.purdue.edu

References

 J. Araus and J. Cairns. Field high-throughput phenotyping: The new crop breeding frontier. *Trends in Plant Science*, 19, October 2013.

- [2] C. Costa, U. Schurr, F. Loreto, P. Menesatti, and S. Carpentier. Plant phenotyping research trends, a science mapping approach. *Frontiers in Plant Science*, 9:1933, 2019.
- [3] A. Habib, W. Xiong, F. He, H. L. Yang, and M. Crawford. Improving orthorectification of uav-based push-broom scanner imagery using derived orthophotos from frame cameras. *IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing*, 10:1–15, 02 2016.
- [4] K. Johansen, M. Morton, Y. Malbeteau, B. Aragon, S. Al-Mashharawi, M. Ziliani, Y. Angel, G. Fiene, S. Negrao, M. Mousa, M. Tester, and M. McCabe. Unmanned aerial vehicle-based phenotyping using morphometric and spectral analysis can quantify responses of wild tomato plants to salinity stress. *Frontiers in Plant Science*, 10, 03 2019.
- [5] Z. Khan and S. Miklavcic. An automatic field plot extraction method from aerial orthomosaic images. *Frontiers in Plant Science*, 10, 05 2019.
- [6] L. Li, Q. Zhang, and D. Huang. A review of imaging techniques for plant phenotyping. *Sensors*, 14(11):20078–20111, 2014.
- [7] Y. Lin, T. Zhou, T. Wang, M. Crawford, and A. Habib. New orthophoto generation strategies from uav and ground remote sensing platforms for high-throughput phenotyping. *Remote Sensing*, 13:860, 02 2021.
- [8] N. Otsu. A threshold selection method from gray-level histograms. *IEEE Transactions on Systems, Man, and Cybernetics*, 9:62–66, Janurary 1979.
- [9] D. Powers. Evaluation: From precision, recall and f-factor to roc, informedness, markedness & correlation. *Journal of Machine Learning Technologies*, 2(1):37–63, 2011.
- [10] J. R. Prat. Image-Based Plant Phenotyping Using Machine Learning. PhD thesis, Purdue University, West Lafayette, IN, May 2019. Available at https://doi.org/10.25394/PGS.7774313.v1.
- [11] Gaurav Sharma. Digital Color Imaging Handbook. CRC Press, Boca Raton, FL, 2003.
- [12] Z. Tang, A. Parajuli, C. J. Chen, Y. Hu, S. Revolinski, C. A. Medina, S. Lin, Z. Zhang, and L. Yu. Validation of uavbased alfalfa biomass predictability using photogrammetry with fully automatic plot segmentation. *Scientific Reports*, 11:3336, 02 2021.
- [13] L. Tresch, Y. Mu, A. Itoh, A. Kaga, K. Taguchi, M. Hirafuji, S. Ninomiya, and W. Guo. Easy mpe: Extraction of quality microplot images for uav-based high-throughput field phenotyping. *Plant Phenomics*, 2019:1–9, 11 2019.
- [14] R. L. Vanderlip. *How a Sorghum Plant Develops*. Kansas State University, Manhattan, KS, 1972.
- [15] T. Wiesner-Hanks, H. Wu, E. Stewart, C. DeChant, N. Kaczmar, H. Lipson, M. Gore, and R. Nelson. Millimeter-level plant disease detection from aerial photographs via deep learning and crowdsourced data. *Frontiers in Plant Science*, 10, 2019.
- [16] C. Xie and C. Yang. A review on plant high-throughput phenotyping traits using uav-based sensors. *Computers and Electronics in Agriculture*, 178:105731, 11 2020.
- [17] K. Yang, S. Chapman, N. Carpenter, G. Hammer, G. McLean, B. Zheng, Y. Chen, E.J. Delp, A. Masjedi, M.

Crawford, et al. Integrating crop growth models with remote sensing for predicting biomass yield of sorghum. *in silico Plants*, 3(1):1, 2021.

[18] W. Yang, H. Feng, X. Zhang, J. Zhang, J. Doonan, W. Batchelor, L. Xiong, and J. Yan. Crop phenomics and highthroughput phenotyping: Past decades, current challenges, and future perspectives. *Molecular Plant*, 13, 01 2020.